

RESEARCH ARTICLE

VITAMIN C PREVENTS STRESS INDUCED CARDIOMYOPATHY IN PRENATAL NOISE EXPOSED RODENTS

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Abstract: Potency of oral vitamin C to prevent cardiomyopathy in prenatal noise exposed newborn Wistar rats was studied by comparing the cardiomyocyte numbers and the extracellular matrix expressions (ECM) to controls. Twenty-four newborns (NR) of 32 pregnant mothers were divided equally into 4 groups: K1 (distilled water [DW]), K2 (150 mg/kg of BW oral vitamin C once daily [VC]), P1 (4 hours daily of white noise at 95 dB [WN]+DW), and P2 (WN+VC). VC and WN were given from D1 till birth and from D15 till birth, respectively. The hearts of NR were harvested, and processed for histology slides (2 midsagittal 4 µ cut slides/ animal) stained with hematoxyllin-eosin and Masson trichrome for the cardiomyocytes and ECM quantification at the ventricles using Image Raster 3.0 and ImageJ, respectively. Pictures from 8 visual fields/ slide were taken and analyzed in duplicate (400x magnifications under a light microscope). Data were analyzed using SPSS 17; significance level of p<0.05. In P1, the cardiomyocite cell numbers was significantly lowest (p<0.001); whilst the ECM was significantly highest than K1, K2, P2 (p<0.001 and p<0.005, respectively). Here, vitamin C could prevent the adverse effect of prenatal noise exposures in the ventricle myocardium of newborn rats.

Keywords: cardiomyopathy, hypertrophy, prenatal noise, vitamin C

INTRODUCTION

Prenatal noise exposures may alter the DNA of cardiomyocytes in the heart of the rodents. At the same time, ultrastructural of the rat cardiomyocytes showed several subcellular changes (1). Noise exposure may increase fibrosis in rat cordis. This noise effect can result in extracellular matrix changes characterized by abnormal proliferation of collagen and thus development of tissue fibrosis (2). The alterations of heart tissue may disrupt the function of this organ correlating to many pathology i.e. hypertrophy cardiomyopathy, hypertension and ischemic heart disease (3).

In newborn rats, administration of vitamin C prevented tissue injury and death. Furthermore, vitamin C can improve cardiac protection during

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myocardial repair (4). Using isolated cardiomyocytes, it was reported that vitamin C reduced the necrosis and apoptosis of the cells (5). The study on the newborn rat ventricles treated with oral vitamin C after prenatal noise exposure has yet been cleared, thus studied here.

METHODS

The experimental protocols had been approved by the Animal Care and Use Committee (ACUC) of The Faculty of Veterinary Medicine, Universitas Airlangga, Surabaya.

Twenty-four newborns from 32 female Wistar rats divided into 4 groups equally: K1 (distilled water), K2 (vitamin C 150 mg/kg BW/day), P1 (distilled water + white noise, 95 dB, 4 hr/day), P2 (vitamin C 150 mg/kg BW/day + white noise, 95 dB, 4 hr/day). Vitamin C and distilled water was administered orally once daily at 9 am from day 1 of pregnancy until delivery (6). Noise was given to P1 and P2 group as a white noise generated from a Realtime analyzer software version 5.2.0 (Yoshimasa Electronic Inc., Japan) that connected to a loudspeaker (Sony SRS XB30, Japan) located 30 cm above the rat cages, from day 15 of pregnancy until delivery (at 10 am to 2 pm daily). The intensity of the noise was measured by a sound level meter (Krisbow, Indonesia). K1 and K2 group were kept in different chambers without noise exposures in a different room during the experiment as control groups. At birth, the newborns were sacrificed (n=6 in each group).

The hearts of the newborn rats were harvested after decapitation, fixed in 10% formaldehyde contained solution. In all series, the hearts were sectioned from the apex cordis of the ventricle. Prior to this, the whole heart was paraffinated and sliced at midsagittaly, midventricular the fragment was selected for histology Slides were stained hematoxylin-eosin for the counting of number of nuclei of cardiomyocytes using Image Raster 3.0 software (Mikonos Transdata Nusantara, Indonesia). The adjacent slide stained using Masson's trichrome to calculate the fibrosis in the ventricle myocardium represented by the % area of the ECM using ImageJ software (NIH, USA) (7). All slides were analyzed under a light microscope (Olympus, Japan) twice independently at 400x of magnification (2 slides/ animal; 8 visual fields/ animal). Data were analyzed either using ANOVA and LSD post-test; or Kruskall-Wallis and Mann-Whitney post-test. Prior to these tests, data were analyzed Saphiro-Wilks using and Levene homogeneity tests. The significance level is p<0.05 (SPSS 17).

RESULTS

The numbers of the cardiomyocytes of all groups were presented in table 1. The % area of the ECM presented of fibrosis area from all groups was detailed in table 2.



Table 1. The results of the number of cardiomyocyte cell heart in each group. The highest count was observed in K2, whilst the lowest in P1.

	Cardiomyocyte				
Groups	Median (Maximum- Minimum)	Kruskal Wallis	Sig. (p) of Mann Whitney post test		
K1 (n=6)	89.25 (94.75 – 79.00)		K2 p=0.004		
			P1 p=0.004		
K2 (n=6)	104.75 (110.56 – 100.75)		P2 p=0.005		
			K1 p=0.004 P1 p=0.004		
			P2 p=0.004		
P1 (n=6)	57.41 (66.75 – 56.31) ^a	$p < 0.001^{a}$	K1 p=0.004*		
			K2 p=0.004		
			P2 p=0.004		
P2 (n=6)	90.47 (93.88 – 78.50)		K1 p=0.005		
			K2 p=0.004		
			P1 p=0.004		

Data are expressed as median number ± standard of error ^a P1 vs. K1, K2, P2. *p<0.05

After normality and homogeneity tests, we found that the data of the cardiomyocte numbers not distributed normally although homogenous. Thus proceeded with Kruskal-Wallis and Mann-Whitney post-test. compared to the other groups, the number of the cardiomyocytes in P1 was significantly lower (p<0.05). In the post-test, it was shown that the cardiomyocyte number in P2 was

significantly higher compared to in P1 (p=0.004).

Prior to the inferential stats, the normality and homogeneity tests were conducted using Shapiro-Wilks and Levene tests. The % area of ECM was distributed homogenous and normally. Thus, the % area of ECM analyzed using **ANOVA** followed by LSD post test. It was found that the fibrosis represented by the % of ECM in P1

Tabel 2. The results of the area of fibrosis heart in each group.

C	Area of fibrosis				
Groups	Rerata ± SE	ANOVA	Sig. (p) LSD post test		
	9886,37 ± 557,88		K2 p<0.0001		
K1 (n=6)			P1 p<0.0001		
			P2 p<0.0001		
	$14450,18 \pm 719,13$		K1 p<0.0001		
K2 (n=6)			P1 p<0.0001		
		0 001 ⁸	P2 p<0.0001		
	$24275,\!38 \pm 1058,\!05$	$p=0.001^{a}$	K1 p<0.0001		
P1 (n=6)			K2 p<0.0001		
			P2 p=0.003		
P2 (n=6)	$20823,83 \pm 476,12$		K1 p<0.0001		
			K2 p<0.0001		
			P1 p=0.003		

Data are expressed as mean \pm standard error of the mean. ^a P1 vs. K1, K2, P2. The area of fibrosis in P1 was significantly highest (p=0.001).



was significantly higher compared to the other groups (p<0.001) showed in table 2. In LSD post-test, it was shown that the fibrosis area in P2 was significantly lower when compared to in P1 (p=0.003; table 2).

The representative figures of myocardium ventricles of 4 groups were shown below (Figure 1). The cardiomyocytes counted from the nucleus of each cell, it is oval and situated centrally, coloured in darker pinkish-purple. The ECM was shown as cotton-candy like structure coloured in lighter pinkish purple.

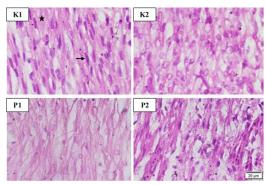


Figure 1. Cardiomyocytes cell in the newborn rat cardiovascular of K1, K2, P1 and P2 at 400x magnification with hematoxyllin-eosin staining. Scale bar 20 μ . Numbers of cardiomyocytes in P1 were decreased significantly compared to other groups (p<0.001). Whilst in P2, the cardiomyocytes were significantly higher compared to in P1 (p<0.05). Arrow showed the nucleus of the cardiomyocytes, the black star showed the ECM.

Distribution area of collagen at ECM represented the potential fibrosis shown in 5 steps of colour deconvolution using ImageJ can be seen in Figure 2. In P1, the area of fibrosis increase compared to groups K1, K2 and P2 which were statistically significant (p = 0.001).

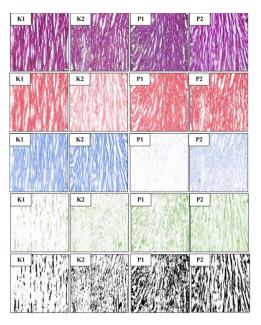


Figure 2. Area of fibrosis in the newborn rat heart of K1, K2, P1 and P2 at 400x magnification with masson trichome staining. Scale bar $20~\mu$. Area of fibrosis in P1 were increased significantly after a prenatal noise-stress exposures compared to other groups (p<0.001).

DISCUSSIONS

We observed significant the cardiomyocyte decrease in numbers in the myocardium of P1 ventricles when compared to P2 and controls. The myocardium is a highly organized structure that contains several cell types, such cardiomyocytes and fibroblasts. At the cellular level, cardiac growth occurs because of cellular proliferation and increased cellular volume (8). A newborn heart contains about half of the total number of myocytes compared to the heart of an adult. The volume of cardiomyocytes in humans, and also in rats, increases almost 25-folds and the number of cardiomyocytes become 3-4 times higher from birth to 2 months (9). The number of cardiomyocytes nucleus is one of the parameters that can be used



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determine the occurrence of cardiomyocytes apoptosis. Previous study reported that noise could be a source of oxidative stress that has an adverse effect to the biological tissue including hearts. Noise can modulate the blood pressure, heart pulse and the levels of stress hormones i.e. glucocorticoids (10). In humans, fetus might response to acoustic external stimulants as early as 2 months; where noise can be transmitted via abdominal walls of the mother thus could affect the fetus directly. The approximate noise threshold was approximately 90-100 dB before producing structural damage to the hearing organs (11).

In oxidative stress, free radicals are atoms and/ molecules that have unpaired electrons in order to obtain chemical stability, immediately bound to the surrounding biology structures. Structures that are picked up by electrons will become free radicals as well so initiate a chain reaction, which eventually causes cell damages (12). The body would response to these stressors by developing general stress syndrome or general adaptation (GAS). syndrome **GAS** would activate the hypothalamushypophysis-adrenal (HPA) axis by releasing the corticotropin-releasing which hormone (CRH) stimulates the adrenocorticotropic hormone (ACTH) secretion into the circulation. This hormone can be transported via the placentae from maternal to fetal circulation (13,14). The increase glucocorticoids may disrupt the gluconeogenesis glycolisis by producing a relative hyperglycemia where glucose would go under pylol pathway or sorbitolaldose reductase pathway, which can be transferred into sorbitol via aldose that transformed reduktase nicotinamide adenine dinucleotide phophatase (NADPH) into NADP. The sorbitol would also be changed into fructose by transformation of NAD+ into NADH. The decrease of NADH could deplete the production of glutathione (GSH) as the natural anti-oxidant thus implicating imbalance between the oxidants and the anti-oxidants resulted in oxidative stress. This condition would then provoke the increase of reactive oxygen species (ROS) (15).

In this study we also found that there was an increase expressions of the ECM, one of potential markers of myocardium fibrosis, in P1 grup compared to in the other groups. On the other hand, the administration of oral vitamin C in P2 could prevent the over deposition of the collagen fibres in the extracellular matrix of the ventricle myocardium stained by Masson Trichrome compared to P1.

may Vitamin C scavengers to free radicals, including ROS, thus stopping the cell damage by producing an ascorbil that lesser reactive. Vitamin C is considered as principal water soluble antioxidant in plasma and after uptake into cells, is able to exert many protective anti-apoptotic and antiinflammatory actions. Vitamin C is a potent endogenous antioxidant crucial for maintaining antioxidant status and scavenging free radicals (16). These of may be the cause the cardiomyocytes lesser degree loss seen in P2 compared to in P1 in the current study.

Furthermore, vitamin C plays an important role for balancing the synthesis and deposition of collagen



protein, which is the most abundant protein in the body and has a strong impact on the composition, structure and biomechanics of extracellular matrix tissue (16). The ECM is a complex mix of fibrillar polysaccharides and proteins synthesized and secreted by the cells in the extracellular space (17). The ECM homeostasis requires a balance degradation between the synthesis of the collagen and, changes in this interaction may result in an abnormal collagen network in the heart (18,19). In the heart, fibroblasts are important cells in the remodeling that occurs in response to pathology changes such as, hypertension, mvocardial infarction and heart failure (19).The clinical consequences of its presence would be inherent to the deposition of collagen in the cardiomyocytes and around the cardiac vessels leading to myocardial stiffness, left ventricular dysfunction, modifications coronary flow reserve and ventricular

CONCLUSIONS

arrhythmias (20).

In this study, vitamin C may prevent stress induced damage on the heart of prenatal noise exposed rodents.

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REFERENCE

- 1. Lenzi P, Frenzilli G, Gesi M, Ferrucci M, Lazzeri G, Fornai F. DNA damage associated with ultrastructural alterations in rat myocardium after loud noise exposure. Environ Health Perspect 2003;111:467–471.
- Alves-Pereira M, Castelo Branco NAA. Vibroacoustic disease: biological effects of infrasound low-frequency noise explained by mechano transduction cellular signaling. **Prog** Bioph Mol Biol. 2007;93:256-279.
- 3. Babish W. Road traffic noise and cardiovascular risk. Noise & Health. 2009;10 27–33.
- 4. Gao F, Yao CL, Gao E, Mo QZ, Yan WL, McLaughlin R, Lopez BL, Christopher TA & Ma XL. Enhancement of glutathione cardioprotection by ascorbic acid in myocardial reperfusion injury. J. Pharmacol. Exp. Ther 2002;301:543–550.
- 5. Akolkar G, Bagchi AK, Ayyappan P, Jassal DS & Singal PK. Doxorubicin-induced nitrosative stress is mitigated by vitamin C via the modulation of nitric oxide synthases. Am J Physiol Cell Physiol 2017;312: C418-C427.
- 6. Ibrahim BS. Barioni ED. Heluany E, Braga TT. Beneficial effects of vitamin C treatment on pregnant exposed rats to formaldehyde: reversal of immunosuppression the Toxicol. offspring Appl. Pharmacol 2016.
- 7. Chen Y, Yu Q, Xu CB. A convenient method for quantifying collagen fibers in



- atherosclerotic lesions by ImageJ software. Int J Clin Exp Med 2017;10(10): 14904-14910.
- 8. Anderson RH, Ho SY, Becker AE. Anatomy of the human atrioventricular junctions revisited. Anat Rec 2000;260:81–91.
- 9. Hew KW, Keller KA. Postnatal anatomical and functional development of the heart: a species comparison. Birth Defects Res. B Dev. Reprod. Toxicol 2003;68:309–320.
- 10. Tsalogli dou A, Koukourikos K, Pantelidou P, Katsimbeli A, Monios A, Kourkouta L. Noise Pollution as a Cardiovascular Health Hazard. International Journal of Engineering and Applied Sciences (IJEAS) 2015; vol.2: no 82-85.
- 11. Porcaro C, Zappasodi F, Barbati G, Salustri C, Pizzella V, Rossini PM, et al. Fetal auditory responses to external sounds and mother's heart beat: Detection improved by Independent Component Analysis. Brain Res 2006:1101:51-8.
- 12. Halliwell B, Whiteman M. Measuring reactive species and oxidative damage in vivo and in cell culture: how should you do it and what do the results mean?. Br J Pharmacol 2004;142:231-55.
- 13. Sherwood L. Human Physiology: From Cells To Systems. In: Human Physilogy. Cengage Learning, Inc, Mason, OH, United States 2010;702-712.
- 14. Shier D, Butler J and Lewis R. Hole's Human Anatomy & Physiology, 14 ed., McGraw-Hill

- Education–Europe, London, United States 2007.
- 15. Rains JL and Jain SK. Oxidative Stress, Insulin Signaling, And Diabetes. Free Radical Biology and Medicine 2011 vol. 50, no. 5:567-575.
- 16. Sagun KC, Carcamo JM, Golde DW. Vitamin C enters mitochondria via facilitative glucose transporter 1 (Glut1) and confers mitochondrial protection against oxidative injury. Faseb J 2005;19:1657–67.
- 17. L. Roll and A. Faissner. Influence of the extracellular matrix on endogenous and transplanted stem cells after brain damage. Frontiers in Cellular Neuroscience 2014;vol. 8, p. 219.
- 18. C. D'Aniello, E. Habibi, F. Cermola et al. Vitamin C and l-proline antagonistic effects capture alternative states in the pluripotency continuum. Stem Cell Reports 2016;vol. 8, no.1, pp. 1–10.
- 19. Van den Borne SWM, Diez J, Blankesteijn M, Verjans J, Hofstra L, Narula J. Myocardial remodeling after infarction: the role of myofibroblasts. Nat Rev Cardiol 2010;7:30–37.
- 20. Barasch E, Gottdiener JS, Aurigemma G, Kitzman DW, Han J, Kop WJ, Tracy RP. Association between elevated fibrosis markers and heart failure in the elderly. The cardiovascular health study. Circ Heart Fail 2009;2:303 310.